



Certificate of Analysis - Amended

Product Description	WA27	
Cell Line Provider	WiCell	
Parent Material	This material descended from derivation	
Lot Number	WB0130	
Date Viald	26-February-2012	
Passage Number	p8 ¹	
Culture Platform	Feeder Independent	
	Medium: E8 plus PVA	Matrix: Recombinant Human Vitronectin

The following testing specifications have been met for the specified product lot.

Test Description	Test Provider	Test Method	Test Specification	Result
Post-Thaw Viable Cell Recovery	WiCell	SOP-CH-305	≥ 15 Undifferentiated Colonies, ≤ 30% Differentiation	Pass ²
Identity by STR	UW Molecular Diagnostics Laboratory	PowerPlex 16 HS System by Promega	Consistent with known profile	Pass
Sterility - Direct transfer method	Apptec	30744	Negative	Pass
Mycoplasma	Charles River	ID 91/0	Negative	Pass
Karyotype by G-banding	WiCell	SOP-CH-003	Normal karyotype	Pass

¹These cells were cultured for 7 passages prior to freeze. WiCell adds +1 to the passage number at freeze so that the number on the vial best represents the overall passage number of the cells at thaw. Footnote provided by TL on 17Sep12.

²Post-Thaw Viable Cell Recovery was obtained using Rho-Kinase Inhibitor (Y-27632) during thaw.

The following tests were performed on the cell line. The tests do not apply to any particular lot.
Please see the individual test reports for results of each test.

Test Description	Test Provider	Test Method
Differentiation Potential by Teratoma	WiCell	SOP-CH-213 SOP-CH-214
HLA	UW Histocompatibility Laboratory	High resolution sequencing method with Celera reagents on the ABI 3100 instrument
ABO	New York Blood Center	For ABO: Olsson ML, Chester MA. A rapid and simple ABO genotype screening method using a novel B/O2 versus A/O2 discriminating nucleotide substitution at the ABO locus. Vox Sang 1995; 69(3):242-7. For RHD: Singleton BK, Green CA, Avent ND, Martin PG, Smart E, Daka A, Narter-Olaga EG, Hawthorne LM, Daniels G. The presence of an RHD pseudogene containing a 37 base pair duplication and a nonsense mutation in Africans with the Rh D-negative blood group phenotype. Blood 2000; 95(1): 12-8.
Growth Curve (Doubling Time)	WiCell	Varies by culture platform
Flow Cytometry for ESC Marker Expression	WiCell	SOP-CH-024
Comprehensive Human Virus Panel	Charles River	ID 91/0



Certificate of Analysis - Amended

Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.

Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells.

Amendment(s):

Reason for Amendment	Date
CoA updated to include copyright information.	See Signature
Amended STR test method and HLA test provider and test method.	05-October-2012
Original CoA.	16-August-2012

Date of Lot Release	Quality Assurance Approval
16-August-2012	1/3/2014 X AMC AMC Quality Assurance Signed by: [REDACTED]



Short Tandem Repeat Analysis*

Sample Report: 10447-STR

Label on Tube: 10447-STR

Sample Date: 04/20/12

Requestor: WiCell Research Institute

Lab Received 04/20/12

Test Date: 04/25/12

File Name: 120425_CLN

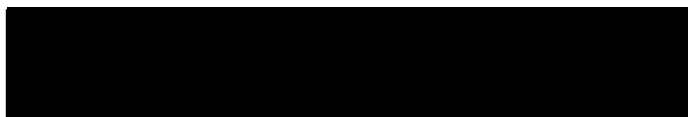
Report Date: 04/30/12

Sample Name: (label on tube) 10447-STR

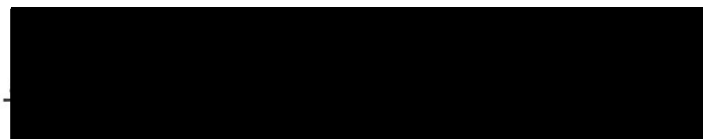
Description: WI Cell Research Institute provided
genomic DNA
236 ug/mL 260/280=1.88

Locus	Repeat #	STR Genotype
D16S539	5, 8-15	12,13
D7S820	6-14	10,10
D13S317	7-15	11,11
D5S818	7-15	11,12
CSF1PO	6-15	11,13
TPOX	6-13	8,10
Amelogenin	NA	X,X
TH01	5-11	6,8
vWA	11, 13-21	16,18

Comments: Based on the 10447-STR DNA submitted by WI Cell dated and received on 04/20/12, this sample (Label on Tube Cap only: 10447-STR) exactly matches the STR profile of the human stem cell line WA27 comprising 14 allelic polymorphisms across the 8 STR loci analyzed. No STR polymorphisms other than those corresponding to the human WA27 stem cell line were detected and the concentration of DNA required to achieve an acceptable STR genotype (signal/ noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. This result suggests that the 10447-STR DNA sample submitted corresponds to the WA27 stem cell line and was not contaminated with any other human stem cells or a significant amount of mouse feeder layer cells. Sensitivity limits for detection of STR polymorphisms unique to either this or other human stem cell lines is ~5%.



Molecular Diagnostics Laboratory



Molecular Diagnostics Laboratory

* Testing to assess engraftment following bone marrow transplantation was accomplished by analysis of human genetic polymorphisms at STR loci. This methodology has not yet been approved by the FDA and is for investigational use only.

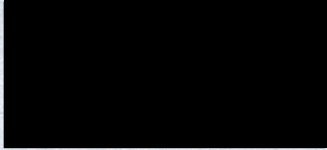
Test Facility:



This report is confidential. No part may be used for advertising or public announcement without written permission. Results apply only to the sample(s) tested.

Report Number
900594
Page 1 of 1

WiCell Research Institute



June 12, 2012
P.O. #:

STERILITY TEST REPORT

Sample Information:

Stem Cells

- 1: WA27-WB0130 10504
- 2: WA26-WB0131 10505
- 3: WA25-WB0132 10506
- 4: WA25-WB0127 10507
- 5: WA26-WB0128 10508
- 6: WA27-WB0138 10509
- 7: WA26-WB0152 10514
- 8: WA25-WB0151 10512
- 9: WA09-WB0143 10521
- 10: WA09-WB0139 10520
- 11: WA27-WB0150 10522
- 12: H9 hOct4-pGZ-WB0140 10518
- 13: MIRJT6i-mND1-4-WB0142.10519

Date Received:

May 23, 2012

Date in Test:

May 29, 2012

Date Completed:

June 12, 2012

Test Information:

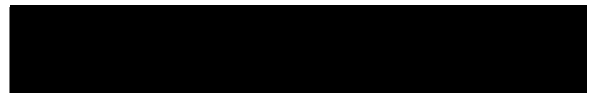
Test Codes: 30744, 30744A
Immersion, USP / 21 CFR 610.12
Procedure #: BS210WCR.201

TEST PARAMETERS	PRODUCT	
Approximate Volume Tested	0.5 mL	0.5 mL
Number Tested	26	26
Type of Media	SCD	FTM
Media Volume	400 mL	400 mL
Incubation Period	14 Days	14 Days
Incubation Temperature	20 °C to 25 °C	30 °C to 35 °C
RESULTS	26 NEGATIVE	26 NEGATIVE



QA Reviewer

Date



Technical Reviewer

Date

Testing conducted in accordance with current Good Manufacturing Practices.



Sponsor: WiCell Research Institute

Accession #: 2012-015912

Diagnostic Summary Report



Received: 20 Mar 2012
Approved: 27 Mar 2012, 13:11
Bill Method:
Test Specimen: Human

Table with 10 columns: Sample Set, Service (# Tested), Profile, Assay, Tested, +, +/-, ?, PDG. Row 1: #1, Infectious Disease PCR (3), All Results Negative.

+ = Positive, +/- = Equivocal, ? = Indeterminate, PDG = Pending

Service Approvals

Table with 3 columns: Service, Approved By*, Date. Row 1: Infectious Disease PCR, [Redacted], 27 Mar 2012, 13:11

To assure the SPF status of your research animal colonies, it is essential that you understand the sources, pathobiology, diagnosis and control of pathogens and other adventitious infectious agents that may cause research interference. We have summarized this important information in infectious agent Technical Sheets, which you can view by visiting http://www.criver.com/info/disease_sheets.

*This report has been electronically signed by laboratory personnel. The name of the individual who approved these results appears in the header of this service report. All services are performed in accordance with and subject to General Terms and Conditions of Sale found in the Charles River Laboratories-Research Models and Services catalogue and on the back of invoices.

Sponsor: WiCell Research Institute

Accession #: 2012-015912

Product: Not Indicated

Test Specimen: Human

Received: 20 Mar 2012

Molecular Diagnostics Infectious Disease PCR Results Report

Department Review: Approved by [REDACTED] 27 Mar 2012, 13:11*

Human Comprehensive Virus Panel

Sample #: Code :	<u>1</u>	<u>2</u>	<u>3</u>
	WA25-WB0132 10429	WA26-WB0131 10430	WA27-WB0130 10431
<i>John Cunningham virus</i>	-	-	-
<i>BK virus</i>	-	-	-
<i>Herpesvirus type 6</i>	-	-	-
<i>Herpesvirus type 7</i>	-	-	-
<i>Herpesvirus type 8</i>	-	-	-
<i>Parvovirus B19</i>	-	-	-
<i>Epstein-Barr Virus</i>	-	-	-
<i>Hepatitis A virus</i>	-	-	-
<i>Hepatitis B virus</i>	-	-	-
<i>Hepatitis C virus</i>	-	-	-
<i>HPV-16</i>	-	-	-
<i>HPV-18</i>	-	-	-
<i>Human T-lymphotropic virus</i>	-	-	-
<i>Human cytomegalovirus</i>	-	-	-
<i>HIV-1</i>	-	-	-
<i>HIV-2</i>	-	-	-
<i>Adeno-associated virus</i>	-	-	-
<i>Human Foamy Virus</i>	-	-	-
<i>LCMV PCR</i>	-	-	-
<i>Hantavirus Hantaan PCR</i>	-	-	-
<i>Hantavirus Seoul PCR</i>	-	-	-
<i>Mycoplasma Genus PCR</i>	-	-	-
<i>DNA Spike</i>	PASS	PASS	PASS
<i>RNA Spike</i>	PASS	PASS	PASS
<i>NRC</i>	PASS	PASS	PASS

Remarks: - = Negative; I = Inhibition, +/- = Equivocal; + = Positive.

Sample Suitability/Detection of PCR Inhibition:

Sample DNA or RNA is spiked with a low-copy number of a exogenous DNA or RNA template respectively. A spike template-specific PCR assay is used to test for the spike template for the purpose of determining the presence of PCR inhibitors. The RNA spike control is also used to evaluate the reverse-transcription of RNA. Amplification of spike template indicates that there is no detectable inhibition and the assay is valid.

NRC:

The nucleic acid recovery control (NRC) is used to evaluate the recovery of DNA/RNA from the nucleic acid isolation process. The test article is spiked with a low-copy number of DNA/RNA template prior to nucleic acid isolation. A template-specific PCR assay is used to detect the DNA/RNA spike.

**This report has been electronically signed by laboratory personnel. The name of the individual who approved these results appears in the header of this service report.*

Report Date: April 24, 2012

Cell Line: WA27-WB0130 10447

Passage #: 10

Date of Sample: 4/13/2012

Date Completed: 4/24/2012

Specimen: hESC on rh Vitronectin

Cell Line Gender: Female

Reason for Testing: post-freeze

Investigator: [REDACTED] WiCell Derivation

Results: 46,XX



Cell: S02-24

Slide: 2-R1 (20) karyotype

Slide Type: Karyotyping

of Cells Counted: 20

of Cells Karyotyped: 4

of Cells Analyzed: 8

Band Level: 400-575

Interpretation:

No clonal abnormalities were detected at the stated band level of resolution.

Completed by [REDACTED] MS, CG(ASCP), on 4/23/2012

Reviewed and interpreted by [REDACTED], PhD, FACMG, on 4/24/2012

A signed copy of this report is available upon request.

Date: _____

Sent By: _____

Sent To: _____

QC Review By: _____

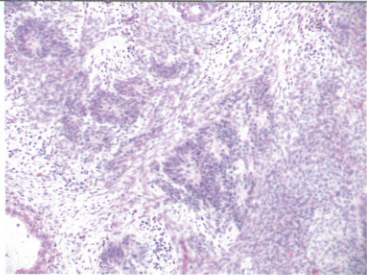
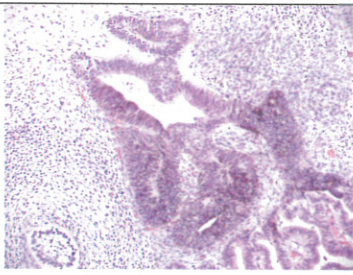
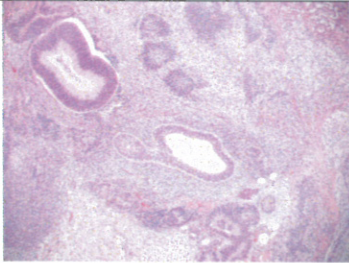
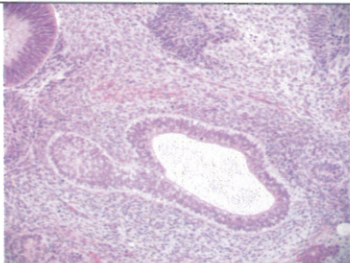
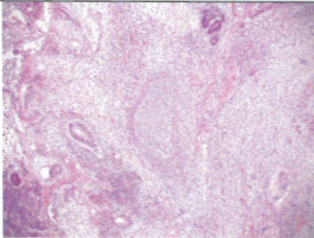
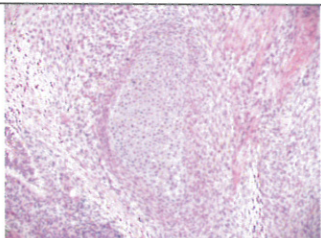
Limitations: This assay allows for microscopic visualization of numerical and structural chromosome abnormalities. The size of structural abnormality that can be detected is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. It is documented here as "band level", i.e., the range of bands determined from the four karyograms in this assay. Detection of heterogeneity of clonal cell populations in this specimen (i.e., mosaicism) is limited by the number of metaphase cells examined, documented here as "# of cells counted".

This assay was conducted solely for listed investigator/institution. The results may not be relied upon by any other party without the prior written consent of the Director of the WiCell Cytogenetics Laboratory. The results of this assay are for research use only. If the results of this assay are to be used for any other purpose, contact the Director of the WiCell Cytogenetics Laboratory.

Cell Line: WA27

Cell Lot Number: NA

Sample Number: 10447-A

ECTODERM	
Structure Name: Neural tubules Magnification:200X Slide ID: A	Structure Name: Pigmented neuroectoderm Magnification: 200X Slide ID: A
	
ENDODERM	
Structure Name: Respiratory Magnification:100X Slide ID: A	Structure Name: Respiratory Magnification:200X Slide ID: A
	
MESODERM	
Structure Name: Cartilage Magnification:100X Slide ID: A	Structure Name: Cartilage 200X Slide ID: A
	

Comments: Structures identified include Ectoderm (2), Mesoderm (1) and Endoderm (1)

Sample(s) were assessed for the presence of differentiation into cell types characteristic of the three embryonic germ layers, which, if present in the sample(s) examined, are represented in the photographs above. The individual's signature below verifies that this report accurately reflects the pathology observed.

Pathologist (By/Date): 6/27/2012

[Redacted Signature]

QA Review (By/Date):

[Redacted Signature] 03/20/12 JTC Error 03/20/12 JTC

© SOP ID SOP-CH-014. Error 03/20/12 JTC

Name: WICELL, 10406_HLA
MRN: OS000184
DOB:
HLA#: WICELL

Hospital:
Physician: ,
Category:

Bone Marrow Case Histocompatibility Summary
301417-DT

HLA Typing Results

Patient	Relation	Hap A*	B*	C*	DRB1*	DRB3*	DRB4*	DRB5*	DQB1*	DPB1*	Tested Date Collect Date
WICELL, 10406_HLA		02:01	41:02	05:01	03:01						03/12/12
OS000184 / WICELL	Patient	66:01	44:02:01G	17:01:01G	04:01						03/01/12

HLA typings performed by sequencing, SSO, SSP or a combination. For low-resolution testing, results are reported by Serologic Equivalents. A "+" in the HLA allele designation indicates that the typing was performed by low/mid-resolution molecular method and that additional alleles are possible. Only the most frequent allele is listed.

HLA DNA-Based Typing

Name HLA / MR# Received	Method Test Date	A*	B*	C*	DRB1*	DRB3*	DRB4*	DRB5*	DQB1*
WICELL, 10406_HLA OS000184 / WICELL 03/01/2012	SEQ 03/20/2012	02:01 66:01							
HLA Allele database: IMGT 3.7.0 2012-01-12									
			41:02						
03/01/2012	SEQ 03/20/2012		44:02:01G						
HLA Allele database: IMGT 3.7.0 2012-01-12 The reported allele group B*44:02:01G includes the following alleles, which share identical sequences in the antigen recognition site of exons 2 and 3: B*44:02 B*44:19N									
				05:01					
03/01/2012	SEQ 03/20/2012			17:01:01G					
HLA Allele database: IMGT 3.7.0 2012-01-12 The reported allele group C*17:01:01G includes the following alleles, which share identical sequences in the antigen recognition site of exons 2 and 3: C*17:01 C*17:02 C*17:03 The following allele combination(s), in which both alleles are listed by the ASHI CWD review committee as rare or not well defined, cannot be excluded: C* 05:29.17:05.									
					03:01				
03/01/2012	SEQ 03/20/2012				04:01				
HLA Allele database: IMGT 3.7.0 2012-01-12 Cannot rule out the rare allele DRB1*03:68N, first identified in August 31, 2011.									

Comments

Name: WICELL, 10406_HLA
MRN: OS000184
DOB:
HLA#: WICELL

Hospital:
Physician: ,
Category:

Bone Marrow Case Histocompatibility Summary
301417-DT

This test was developed and its performance characteristics determined by this laboratory. It has not been cleared or approved by the U.S. Food and Drug Administration. The FDA has determined that such clearance or approval is not necessary. This test is used for clinical purposes. It should not be regarded as investigational or for research. This laboratory is certified under the Clinical Laboratory Improvement Amendments of 1988 (CLIA-88) as qualified to perform high complexity clinical laboratory testing.

Electronically signed by
Kathleen J. Meuer, MT(ASCP),
CHT(A R H I)
Director or Delegate, HLA Laboratory

03/25/2012 12:18
Date/Time



Histocompatibility Laboratory, Room D4/231, 600 Highland Ave., Madison, WI 53792-2472
Teresa Darcy, MD, Medical Director :: Thomas M. Ellis, PhD, D(ABHI) Laboratory Director
Lab: 608.263.8815 (option 3); Fax: 608.263.9610
ASHI: 01-4-WI-03-2, CLIA: 52DO661997

March 20, 2012

WiCell Research Institute
 Attn: Quality Assurance

SAMPLE: DNA WA27 #10406 (MA#168-12)

Date Received: 03/08/12
 Sample Date: 03/01/12

HISTORY: DNA from cell line.

TESTING REQUESTED: Genotype for *ABO* and common *RH*

TESTING PERFORMED: *ABO*: Polymerase chain reaction-restriction fragment length polymorphism (PCR-RFLP) testing for nucleotide (nt) positions 261 (O¹), 467 (A²), 703 (B), and 1096 (B and O²). *RH*: Multiplex PCR-RFLP for *RHD* and *RHCE**C/c. PCR-RFLP for *RHCE* Exon 5 (676C>G for E/e).

DNA MOLECULAR RESULTS: *ABO*: PCR-RFLP testing indicates the presence of a nt261 deleted G, characteristic of O¹ alleles, and the presence of nt703A and nt1096A, characteristic of B alleles. *RH*: *RHD* exons 4 and 7 are present. Negative for the inactivating *RHD* pseudogene. *RH**Cc and *RH**ee.

	<u>Genotype</u>	<u>Predicted Phenotype</u>
WA27 #10406:	<i>ABO</i> *BO ¹ ; <i>RH</i> *D, <i>RH</i> *Cc, <i>RH</i> *ee	<u>Group B; RhD+, C+E-c+e+</u>



Manager, Genomics



Director of Immunohematology and Genomics

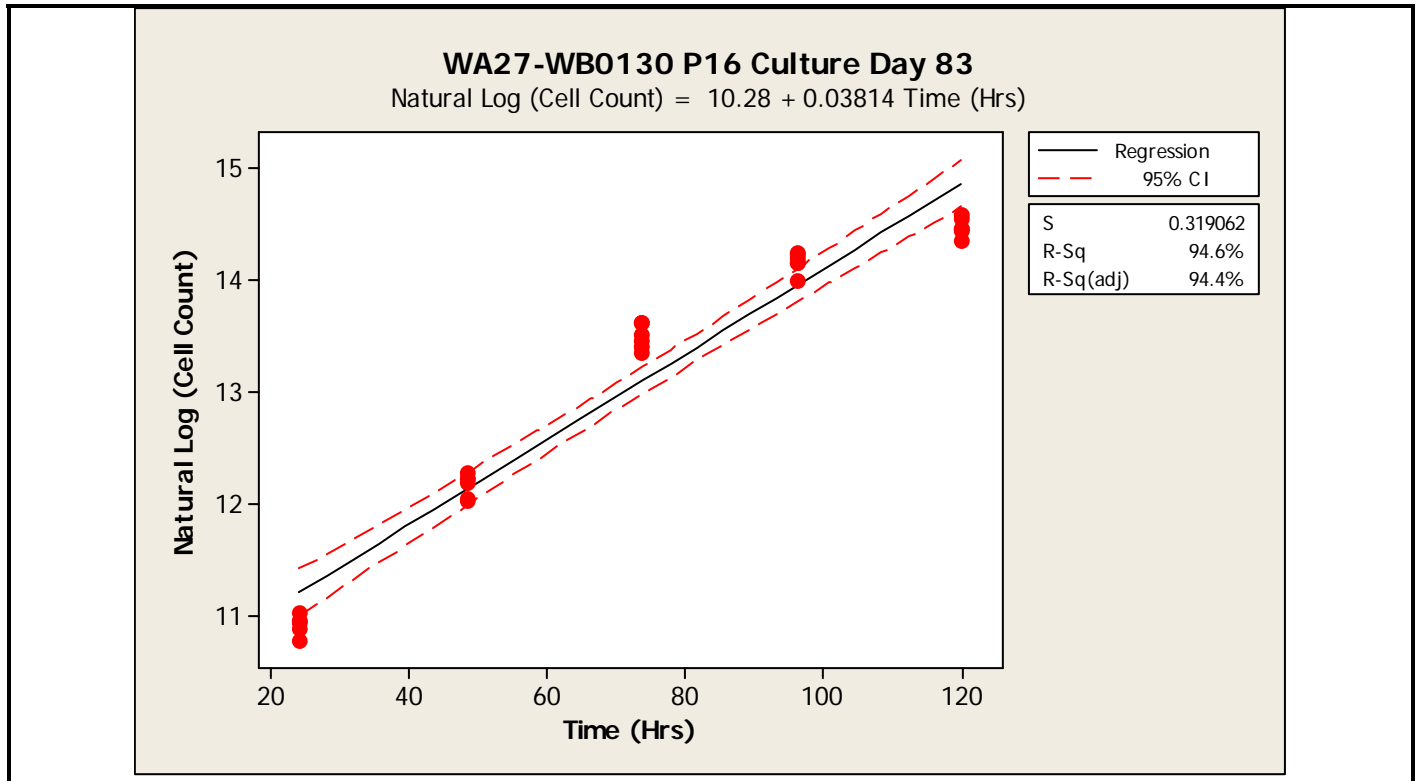
These *in vitro* diagnostic tests were developed and their performance characteristics established in the Molecular Analysis Laboratory. The tests have not been submitted to the Food and Drug Administration (FDA) for clearance or approval and; therefore, are not FDA-licensed tests. The Molecular Analysis Laboratory is certified under the Clinical Laboratory Improvement Amendment (CLIA) of 1988 as qualified to perform high complexity clinical testing. The New York Blood Center has been approved by the New York State Department of Health to perform these tests under its current Clinical Laboratory Permit.

These results are intended to predict a blood group antigen profile in a patient or donor, and are not intended for clinical diagnosis or as the sole means for patient management decisions. There are situations where testing DNA of a person may not reflect the red cell phenotype and not all performance characteristics have been determined. Nucleotide changes that inactivate gene expression or rare new variant alleles may not be identified in these assays. In addition, test results obtained from DNA isolated from leucocytes and other hematopoietic cells may differ from DNA isolated from other tissues in persons with a history of transplantation.



Characterization Report- Growth Characteristics

Sample ID	Cell Line	Cell lot #	Passage	Culture Day	Medium	Matrix	Passaging Additive
10450	WA27	WB0130	16	83	E8 + PVA	rh-Vitronectin	Rho-kinase Inhibitor Y-27632
Documentation of Growth Assay Data				Notebook #	Page(s)	Date Initiated	
				148	73-81	02MAY12	
Growth Assay Performed by		Report Prepared by		Date	QA Reviewed by		Date
WiCell Derivation Laboratory		LAN		14AUG12	JKT		15Aug12



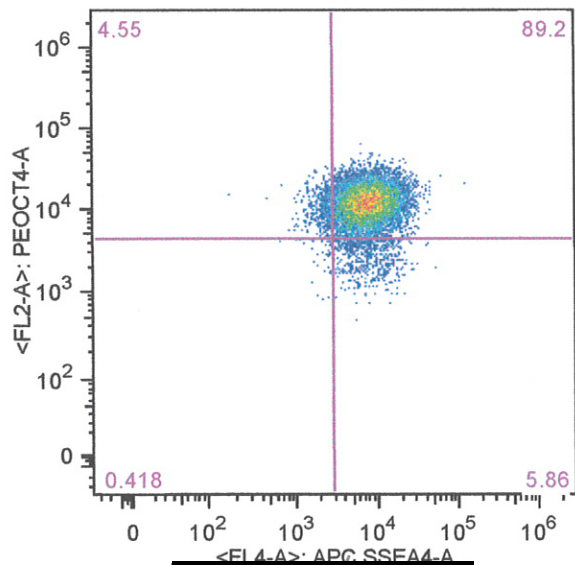
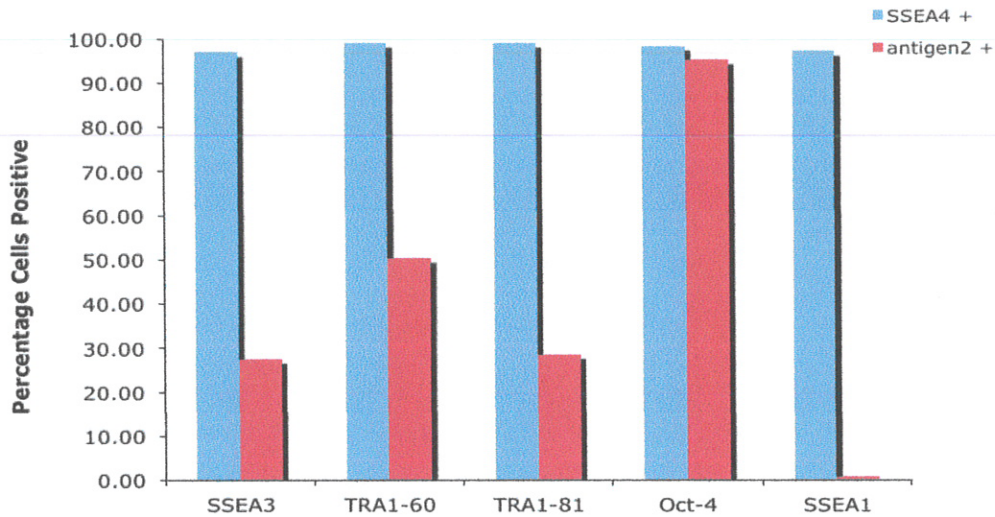
<p style="text-align: center;">Regression Analysis: Natural Log (Cell Count) versus Time (Hrs)</p> <p>The regression equation is Natural Log (Cell Count) = 10.3 + 0.0381 Time (Hrs)</p> <table border="1"> <thead> <tr> <th>Predictor</th> <th>Coef</th> <th>SE Coef</th> <th>T</th> <th>P</th> </tr> </thead> <tbody> <tr> <td>Constant</td> <td>10.2842</td> <td>0.1371</td> <td>75.00</td> <td>0.000</td> </tr> <tr> <td>Time (Hrs)</td> <td>0.038136</td> <td>0.001715</td> <td>22.24</td> <td>0.000</td> </tr> </tbody> </table> <p>S = 0.319062 R-Sq = 94.6% R-Sq(adj) = 94.4%</p> <table border="1"> <thead> <tr> <th colspan="6">Analysis of Variance</th> </tr> <tr> <th>Source</th> <th>DF</th> <th>SS</th> <th>MS</th> <th>F</th> <th>P</th> </tr> </thead> <tbody> <tr> <td>Regression</td> <td>1</td> <td>50.331</td> <td>50.331</td> <td>494.41</td> <td>0.000</td> </tr> <tr> <td>Residual Error</td> <td>28</td> <td>2.850</td> <td>0.102</td> <td></td> <td></td> </tr> <tr> <td>Total</td> <td>29</td> <td>53.181</td> <td></td> <td></td> <td></td> </tr> </tbody> </table>	Predictor	Coef	SE Coef	T	P	Constant	10.2842	0.1371	75.00	0.000	Time (Hrs)	0.038136	0.001715	22.24	0.000	Analysis of Variance						Source	DF	SS	MS	F	P	Regression	1	50.331	50.331	494.41	0.000	Residual Error	28	2.850	0.102			Total	29	53.181				<p>Slope ± 95% C.I. 0.0381 ± 0.0035</p> <p>Apparent Doubling Time (hours) ± 95% C.I. 18.18 ± 2.05</p> <p>Apparent Doubling Time (95% C.I.) 16.64 hours – 20.02 hours</p>
Predictor	Coef	SE Coef	T	P																																										
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Regression	1	50.331	50.331	494.41	0.000																																									
Residual Error	28	2.850	0.102																																											
Total	29	53.181																																												

antigen2:	PERCENTS				ALL SSEA4 +	ALL antigen2 +
	SSEA4 - antigen2 +	SSEA4 + antigen2 +	SSEA4 + antigen2 -	SSEA4 - antigen2 -		
SSEA3	0.37	10.30	78.80	10.50	89.10	10.67
TRA1-60	3.00	64.00	30.60	2.36	94.60	67.00
TRA1-81	2.26	44.00	50.00	3.79	94.00	46.26
Oct-4	4.57	89.2	5.85	0.42	95.05	93.77
SSEA1	0.31	3.08	89.10	7.51	92.18	3.39

Percent analyzable events: 27.9

#wells submitted: 6

Total # cells analyzed: 10.7 X 10⁶



prepared by
 10450



(signature)